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Material-structure-property Optimization and In-process Monitoring of 3D Printed Bone Tissue

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Defect-free 3D printing of biological constructs.

Objectives

- 1. Optimization and testing of the bioprinted constructs in the context of extrusionbased printing of poly(caprolactone) (PCL)-hydroxyapatite (HAp) composite biological constructs.
- 2. Unveil the process- mechanical property relationships in printing of various PCL-HAp composites.
- 3. Demonstrate the use of using in-situ image-based monitoring for quantification of key quality-related aspects of biological constructs, such as geometric integrity.
- 4. Ascertain the *in vitro* osteoinductivity of the fabricated constructs.

Process Parameters \rightarrow In-Situ Sensor Data \rightarrow Construct Quality \rightarrow Mechanical Properties \rightarrow Cell Viability. 4

The Big Picture











Outline

- Background and Rationale
- Methods and Results
 - Optimizing Bioprinting Conditions
 - Mechanical Testing
 - Cell Viability
- Conclusions and Future Work

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Rationale

- Poly(caprolactone) (PCL) is the most popular biopolymer for bone tissue engineering
 - FDA-approved thermoplastic polymer
 - Slow degradation rate (ranging from months to a few years), which is comparable to the rate of bone regeneration.
- PCL is not osteoinductive (bone does not penetrate and grow into the polymer)
 - Hence, mixed with bioceramics and bioglasses
 - Scaffolds of PCL and Hydroxyapatite (Hap) have been successfully 3D printed and tested to induce osteogenesis in vitro and in vivo.

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EnvisionTec Bioprinter with Onboard Camera



Evolutionary Optimization Experimental Methodology

There a large number of printing variables.

Not possible to do a conventional factorial design of experiment.



Linear Speed Optimization Using image analysis of single strands









Every material composition has its own "sweet spot"

Need to understand and explain the rheology of the material.



Finding the sweet spot requires trial-and-error in the 15 absence of a model-based approach.

		Condition 1	Condition 2	Condition 3	Condition 4
		110°C, 1.5 bar	120°C, 2.5 bar	130°C, 3.5 bar	140°C, 4.5 bar
Composition: %PCL/HAp	90/10	8 mm/s	21 mm/s	35 mm/s	>40 mm/s
	80/20	<5 mm/s	9 mm/s	17 mm/s	37 mm/s
	70/30	<5 mm/s	<5 mm/s	9 mm/s	13 mm/s

Printing single strands is trivially simple.

Geometric integrity is important.





Increasing temperature and pressure aids deposition (viscosity decreases) upto a certain threshold

The Circle-Square-Diamond Test



Not all conditions are conducive to print (there is a sweet spot)



The Circle-Square-Diamond Test



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Circularity is used to quantify geometric integrity

$$C = (4\pi A) / P^2$$

C is circularity (dimensionless) A is the area (mm²), and P is perimeter (mm).





Ten circularity measurements per shape and condition

Avoid printing circular scaffolds.



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Compressive testing of solid 10 mm x 10 mm x2.5 mm samples was conducted.

- Four samples were prepared with theoretically solid box architectures for both the lowest and highest print condition and optimal linear print speeds for a composition.
- Compress to a strain of 8% over the course of a minute.
- Compressive modulus was determined as the slope of the linear region of the stress-strain curve.

Higher printing temperature and pressure relates to higher compressive strength.





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Higher printing temperature and pressure relates to higher compressive strength.



Explanation of Poor Compression Results (In-situ Images of the worst performing sample from printing)



Defects are the root cause of poor mechanical performance.





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Explanation of Poor Compression Results (Pictures of the worst performing sample from printing)



Defects are the root cause of poor mechanical performance.



Rheology Testing

To assess the rheological properties of the developed composites during the extrusion process, a rotational rheometer was used to perform routine oscillatory experiments.







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Cell Viability

Effectiveness of the engineered scaffolds in bone tissue engineering is demonstrated by in vitro culture of human mesenchymal stem cells (hMSCs).



Higher HAp concentration correlates to enhanced cell 34 attachment and proliferation.

The viability and proliferation rate of hMSCs were quantified by measuring their metabolic activities using PrestoBlue Cell Viability Reagent on days 1, 3, and 7



70/30 suggests the positive role of HAp in supporting cell attachment and proliferation.

Immunostaining

Cellular expression of osteopontin (OPN) and bone sialoprotein (BSP) after 28 days of seeding confirmed the osteogenic maturation of hMSCs



Immunostaining results demonstrates the viability of 3D printed bone tissues



Both OPN and BSP were present in printed samples.

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Differentiated cells on samples significantly increased from PCL100/H0 to PCL70/H30 samples.



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Xylenol Orange Staining

- Osteodifferentiated cells in the late stage of their differentiation have a high calcium deposition.
- To investigate the functional mineralization of differentiated cells, xylenol orange staining was performed on seeded constructs on day 35 of post differentiation.



Calcification area was significantly increased by increasing the HA percentage





Statistically significant increase in calcification with increase in HAp

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Conclusions

- Higher concentrations of HAp increases osteoinductivity.
- Defects influence properties (hence in-situ monitoring is key)
- Need to model and explain the process-structure relationships.